

**Order information**

REF	CONTENT	Analyzer(s) on which <b>cobas c</b> pack(s) can be used
05973058 190	Tina-quant IgA CSF 150 tests	System-ID 07 7480 4 Roche/Hitachi <b>cobas c</b> 311, <b>cobas c</b> 501/502
06533850 190	Calibrator f.a.s. IgA/IgM CSF (3 x 1 mL)	Code 408
05117003 190	PreciControl ClinChem Multi 1 (20 x 5 mL)	Code 391
05947626 190	PreciControl ClinChem Multi 1 (4 x 5 mL)	Code 391
05117216 190	PreciControl ClinChem Multi 2 (20 x 5 mL)	Code 392
05947774 190	PreciControl ClinChem Multi 2 (4 x 5 mL)	Code 392
04489357 190	Diluent NaCl 9 % (50 mL)	System-ID 07 6869 3

**English****System information**For **cobas c** 311/501 analyzers:**IGA-C**: ACN 436For **cobas c** 502 analyzer:**IGA-C**: ACN 8436**Intended use**

In vitro test for the quantitative determination of IgA specifically in human cerebrospinal fluid and corresponding human serum / plasma on Roche/Hitachi **cobas c** systems.

**Summary**<sup>1,2,3,4,5</sup>

Cerebrospinal fluid (CSF) analysis is a basic tool for diagnosis of neurological diseases.

The diffusion of proteins through the blood-brain barrier normally occurs at a steady rate. The rate is influenced by the permeability of the blood-brain barrier and CSF flow rate. Changes in protein concentration in the CSF can be an indication for various neurological diseases.

Disease-related immunoglobulin patterns (IgG, IgA, IgM with reference to albumin) allow for the differential diagnosis of neurological disorders with the aid of Reiber quotient schemes.

Elevated levels of IgA in CSF are often associated with opportunistic infections of the central nervous system (CNS) and neurotuberculosis. Increased CSF IgA concentrations may occur because of either increased permeability of the blood-brain barrier or local/intrathecal production of IgA, or both. Malfunction of the blood-brain barrier can be reliably quantified by means of the albumin CSF/serum ratio.

Albumin is an ideal reference protein for blood-brain barrier function, since it is solely synthesized outside the brain and thereby provides an excellent measure for proteins passing the blood-brain barrier. An elevated albumin CSF/serum ratio is an indication of disorders of the blood-brain barrier. Measuring IgA and albumin in CSF/serum pairs, a differentiation between IgA originating from blood and IgA originating from intrathecal production is possible.

The results of the CSF/serum ratio for IgA and Albumin, in conjunction with Reiber quotient scheme provide an aid in the diagnosis of functional blood-brain barriers disorders and/or intrathecal IgA synthesis.

In serum, IgA exists in monomeric, dimeric and polymeric forms, of which is about 90 % monomeric, but subject to change in some diseases. In CSF, the monomeric form is also predominant with changes in disease. In bodily secretions, such as saliva, sweat, mucosa and colostrum, IgA exists predominantly in dimeric form in association with an additional protein chain (secretory component).

**Test principle**

Particle-enhanced immunoturbidimetric assay

Human IgA agglutinates with latex particles coated with polyclonal anti-human IgA antibodies. The precipitate is determined turbidimetrically.

**Reagents - working solutions**

**R1** TRIS buffer: 500 mmol/L, pH 8.5; NaCl: 1000 mmol/L; rabbit globulin: 0.10 %; detergents; preservative

**R2** Latex particles coated with anti-human IgA antibodies (rabbit) in TRIS buffer: 10 mmol/L, pH 8.5; stabilizer; preservative

**R3** Reagent for antigen excess check. IgA in diluted serum (human); NaCl: 150 mmol/L; phosphate buffer: 50 mmol/L, pH 7.0; preservative

R1 is in position A, R2 is in position B and R3 is in position C.

**Precautions and warnings**

For in vitro diagnostic use.

Exercise the normal precautions required for handling all laboratory reagents.

Disposal of all waste material should be in accordance with local guidelines. Safety data sheet available for professional user on request.

All human material should be considered potentially infectious. All products derived from human blood are prepared exclusively from the blood of donors tested individually and shown to be free from HBsAg and antibodies to HCV and HIV.

The testing methods applied were FDA-approved or cleared in compliance with the European Directive 98/79/EC, Annex II, List A.

However, as no testing method can rule out the potential risk of infection with absolute certainty, the material should be handled with the same level of care as a patient specimen. In the event of exposure, the directives of the responsible health authorities should be followed.<sup>6,7</sup>

**Reagent handling**

Ready for use

Carefully invert reagent container several times prior to use to ensure that the reagent components are mixed.

**Storage and stability****IGA-C**

Shelf life at 2-8 °C:

See expiration date on **cobas c** pack label.

On-board in use and refrigerated on the analyzer:

12 weeks

**Diluent NaCl 9 %**

Shelf life at 2-8 °C:

See expiration date on **cobas c** pack label.

On-board in use and refrigerated on the analyzer:

12 weeks

**Specimen collection and preparation**

Pairs of CSF/serum or CSF/plasma should be collected at the same time.

For specimen collection and preparation only use suitable tubes or collection containers.

Only the specimens listed below were tested and found acceptable. Cerebrospinal fluid (CSF).

Serum.

Plasma: Li-heparin and K<sub>2</sub>-EDTA plasma

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay.

**Serum and plasma**

<b>Stability:</b> <sup>8</sup>	8 months at 15-25 °C
	8 months at 2-8 °C
	8 months at (-15)-(-25) °C

**CSF**

Samples should be as fresh as possible. Centrifuge samples containing particles and/or cells before performing the assay.

<b>Stability:</b> <sup>8</sup>	1 day at 15-25 °C
	7 days at 2-8 °C
	Storage at (-15)-(-25) °C is not recommended.

**Materials provided**

See "Reagents – working solutions" section for reagents.

**Materials required (but not provided)**

- See "Order information" section
- General laboratory equipment

**Assay**

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

The performance of applications not validated by Roche is not warranted and must be defined by the user.

**Application for sample type CSF****cobas c 311 test definition**

Assay type	2-Point End		
Reaction time / Assay points	10 / 7-23		
Wavelength (sub/main)	800/570 nm		
Reaction direction	Increase		
Units	mg/L		
Reagent pipetting		Diluent (H <sub>2</sub> O)	
R1	120 µL	–	
R2	40 µL	–	
R3	20 µL	–	
<b>Sample volumes</b>	<b>Sample</b>	<b>Sample dilution</b>	
		<b>Sample</b>	<b>Diluent (NaCl)</b>
Normal	8 µL	–	–
Decreased	8 µL	25 µL	75 µL
Increased	16 µL	–	–

**cobas c 501/502 test definition**

Assay type	2-Point End		
Reaction time / Assay points	10 / 13-34		
Wavelength (sub/main)	800/570 nm		
Reaction direction	Increase		
Units	mg/L		
Reagent pipetting		Diluent (H <sub>2</sub> O)	
R1	120 µL	–	
R2	40 µL	–	
R3	20 µL	–	
<b>Sample volumes</b>	<b>Sample</b>	<b>Sample dilution</b>	

Normal	8 µL	–	–
Decreased	8 µL	25 µL	75 µL
Increased	16 µL	–	–

**Application for sample type serum and plasma****cobas c 311 test definition**

Assay type	2-Point End		
Reaction time / Assay points	10 / 7-23		
Wavelength (sub/main)	800/570 nm		
Reaction direction	Increase		
Units	mg/L		
Reagent pipetting		Diluent (H <sub>2</sub> O)	
R1	120 µL	–	
R2	40 µL	–	
R3	20 µL	–	
<b>Sample volumes</b>	<b>Sample</b>	<b>Sample dilution</b>	
		<b>Sample</b>	<b>Diluent (NaCl)</b>
Normal	3 µL	2 µL	178 µL
Decreased	2 µL	1.5 µL	180 µL
Increased	6 µL	2 µL	178 µL

**cobas c 501/502 test definition**

Assay type	2-Point End		
Reaction time / Assay points	10 / 13-34		
Wavelength (sub/main)	800/570 nm		
Reaction direction	Increase		
Units	mg/L		
Reagent pipetting		Diluent (H <sub>2</sub> O)	
R1	120 µL	–	
R2	40 µL	–	
R3	20 µL	–	
<b>Sample volumes</b>	<b>Sample</b>	<b>Sample dilution</b>	
		<b>Sample</b>	<b>Diluent (NaCl)</b>
Normal	3 µL	2 µL	178 µL
Decreased	2 µL	1.5 µL	180 µL
Increased	6 µL	2 µL	178 µL

**Calibration**

Calibrators	S1: H <sub>2</sub> O	
	S2-S6: C.f.a.s. IgA/IgM CSF	
	Multiply the lot-specific C.f.a.s. IgA/IgM CSF calibrator value by the factors below to determine the standard concentrations for the 6-point calibration curve:	
<b>cobas c 311</b>	S2: 0.01955	S5: 0.1564
	S3: 0.05214	S6: 0.2085
	S4: 0.08689	
<b>cobas c 501/502</b>	S2: 0.02084	S5: 0.1667
	S3: 0.05555	S6: 0.2222
	S4: 0.09259	

Calibration mode	RCM2
Calibration frequency	Full calibration
	- after reagent lot change
	- as required following quality control procedures

Traceability: This method has been standardized against the reference preparation of the IRMM (Institute for Reference Materials and Measurements) ERM-DA 470K.<sup>9</sup>

**Quality control**

For quality control, use control materials as listed in the "Order information" section.

In addition, other suitable control material can be used.

CSF:	commercially available control materials
Serum/plasma:	PreciControl ClinChem Multi 1, PreciControl ClinChem Multi 2

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

**Calculation**

Roche/Hitachi **cobas c** systems automatically calculate the analyte concentration of each CSF sample.

To calculate serum/plasma samples in g/L a calculated test must be programmed under *Utility > Calculated Test* on the Roche/Hitachi **cobas c** 311 analyzer and the Roche/Hitachi **cobas c** 501 analyzer. Please use the following settings.

**cobas c 311**

Sample Type	Ser/Pl
Unit of Measure	g/L
Report Name	IgA Serum
Item	IGAS
Formula	IGA-C/932.6

**cobas c 501**

Sample Type	Ser/Pl
Unit of Measure	g/L
Report Name	IgA Serum
Item	IGAS
Formula	IGA-C/1000

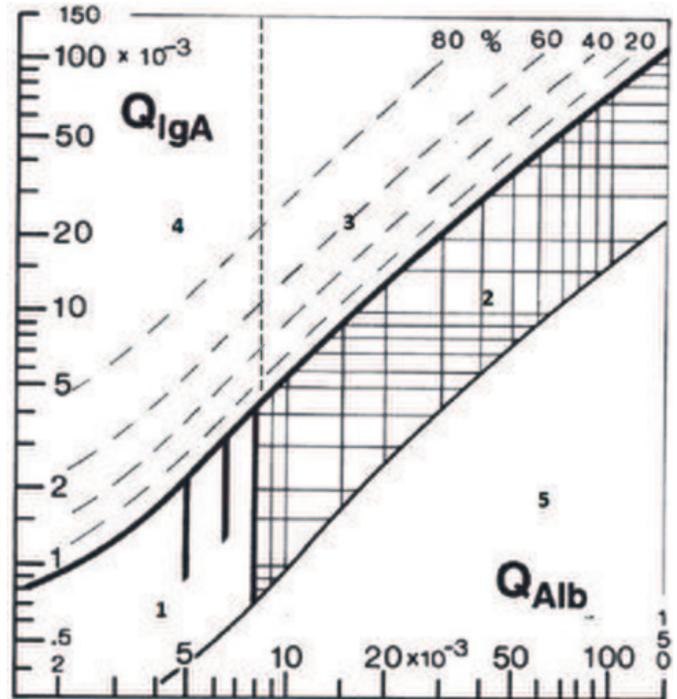
The values for serum/plasma in g/L will be automatically calculated after result output. It is recommended to report the IgA values in serum/plasma to two decimal places, which can be entered in the editable field "Expected Values".

For the definition of the calculated test on **cobas c** 502, refer to the operator's manual of the **cobas** 8000 Data Manager.

**Reiber Quotient Graph**

With the aid of commercially available software, Reiber Quotient Diagrams can be automatically generated.

The calculation employs a ratio diagram including hyperbolic functions as differential lines according to Reiber and Felgenhauer. Results from the determination of IgA and albumin in CSF and serum (IgA and albumin ratios)<sup>10</sup> are plotted.



1. Reference range. 2. Blood brain barrier functional disorder without local IgA synthesis. 3. Blood brain barrier functional disorder with concomitant IgA-synthesis in the CNS. 4. IgA synthesis in the CNS without blood brain barrier disorder. 5. As confirmed empirically, there are no values in this region (i.e. values here are due to errors introduced by blood sampling or analytical errors). Generally speaking, cases not associated with local IgA synthesis in the CNS lie below the bold line (hyperbolic function). The percentage values indicate what percentage of the total IgA in CSF (minimum) originates from IgA synthesis in the CNS relative to the statistically-defined 0 % differential lines (bold).

**Limitations - interference****CSF:**

Criterion: Recovery within  $\pm 10\%$  of initial value.

Icterus: No significant interference up to a conjugated bilirubin concentration of 257  $\mu\text{mol/L}$  or 15 mg/dL.

Hemolysis: No significant interference up to a hemoglobin concentration of 3.11  $\mu\text{mol/L}$  or 5 mg/dL.

High dose hook-effect: Using the prozone check, no false result without a flag was observed up to an IgA concentration of 3000 mg/L.

There is no cross-reaction between IgA and IgG or IgM under the assay conditions.

**Serum/plasma:**

Criterion: Recovery within  $\pm 10\%$  of initial value.

Icterus:<sup>11</sup> No significant interference up to an I index of 60 for conjugated and unconjugated bilirubin (approximate conjugated and unconjugated bilirubin concentration: 1026  $\mu\text{mol/L}$  or 60 mg/dL).

Hemolysis:<sup>11</sup> No significant interference up to an H index of 1000 (approximate hemoglobin concentration: 621  $\mu\text{mol/L}$  or 1000 mg/dL).

Lipemia (Intralipid):<sup>11</sup> No significant interference up to an L index of 2000. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

Rheumatoid factors < 600 IU/mL do not interfere.

High dose hook-effect: Using the prozone check, no false result without a flag was observed up to an IgA concentration of 100 g/L.

There is no cross-reaction between IgA and IgG or IgM under the assay conditions.

Drugs: No interference was found at therapeutic concentrations using common drug panels.<sup>12,13</sup>

As with other turbidimetric or nephelometric procedures, this test may not provide accurate results in patients with monoclonal gammopathy, due to

individual sample characteristics which can be assessed by electrophoresis.<sup>14</sup>

The assay was designed for the determination of IgA in serum/CSF or plasma/CSF pairs only. This assay shall not be used to determine IgA in serum or plasma alone, but always in combination with the matching CSF samples.

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

**ACTION REQUIRED**

**Special Wash Programming:** The use of special wash steps is mandatory when certain test combinations are run together on Roche/Hitachi **cobas c** systems. The latest version of the carry-over evasion list can be found with the NaOH/SMS/Multiclean/SCCS or the NaOH/SMS/SmpCln1+2/SCCS Method Sheets. For further instructions refer to the operator's manual. **cobas c** 502 analyzer: All special wash programming necessary for avoiding carry-over is available via the **cobas** link, manual input is not required.

**Where required, special wash/carry-over evasion programming must be implemented prior to reporting results with this test.**

**Limits and ranges****Measuring range****CSF**

0.4-25 mg/L

Determine samples having higher concentrations via the rerun function. Dilution of samples via the rerun function is a 1:4 dilution. Results from samples diluted using the rerun function are automatically multiplied by a factor of 4.

Determine samples having lower concentrations via the rerun function. For samples with lower concentrations, the rerun function increases the sample volume by a factor of 2. The results are automatically divided by this factor.

**Serum/plasma:**

0.1-6 g/L

Determine samples having higher concentrations via the rerun function. Dilution of samples via the rerun function is a 1:2.2 dilution. Results from samples diluted using the rerun function are automatically multiplied by a factor of 2.2.

Determine samples having lower concentrations via the rerun function. For samples with lower concentrations, the rerun function increases the sample volume by a factor of 2. The results are automatically divided by this factor.

**Lower limits of measurement**

*Limit of Blank (LoB), Limit of Detection (LoD) and Limit of Quantitation (LoQ)*

**CSF:**

Limit of Blank = 0.2 mg/L

Limit of Detection = 0.4 mg/L

Limit of Quantitation = 1.0 mg/L

**Serum/plasma:**

Limit of Blank = 0.05 g/L

Limit of Detection = 0.1 g/L

Limit of Quantitation = 0.25 g/L

The Limit of Blank, Limit of Detection and the Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A2 requirements.

The Limit of Blank is the 95<sup>th</sup> percentile value from n ≥ 60 measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples.

The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

Values below the Limit of Detection (< 0.4 mg/L for CSF and < 0.1 g/L for serum/plasma) will not be flagged by the instrument.

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with a total error of 20 %. It has been determined using low concentration IgA samples.

**Expected values****CSF<sup>15</sup>**

1-3 mg/L

These values are only for orientation. The only relevant values are the CSF/serum ratios.

**Serum/plasma**

Reference values according to CRM 470 Protein Standardization:<sup>16,17</sup>

Adults	0.7-4 g/L
Children and juveniles	
0-1 year	0.00-0.83 g/L
1-3 years	0.20-1.00 g/L
4-6 years	0.27-1.95 g/L
7-9 years	0.34-3.05 g/L
10-11 years	0.53-2.04 g/L
12-13 years	0.58-3.58 g/L
14-15 years	0.47-2.49 g/L
16-19 years	0.61-3.48 g/L

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

**Specific performance data**

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

**Precision**

Repeatability and intermediate precision were determined using human samples and controls in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP5 requirements (2 aliquots per run, 2 runs per day, 21 days). The following results were obtained:

**CSF:**

<i>Repeatability</i>	<i>Mean</i>	<i>SD</i>	<i>CV</i>
	<i>mg/L</i>	<i>mg/L</i>	<i>%</i>
Control 1	8.86	0.07	0.8
Control 2	23.3	0.2	0.7
CSF 1	0.865	0.027	3.1
CSF 2	5.35	0.07	1.4
CSF 3	14.1	0.1	1.0
CSF 4	24.2	0.3	1.1

<i>Intermediate precision</i>	<i>Mean</i>	<i>SD</i>	<i>CV</i>
	<i>mg/L</i>	<i>mg/L</i>	<i>%</i>
Control 1	8.86	0.10	1.1
Control 2	23.3	0.2	1.0
CSF 1	0.865	0.037	4.3
CSF 2	5.35	0.14	2.5
CSF 3	14.1	0.3	1.9
CSF 4	24.2	0.4	1.8

**Serum/plasma:**

<i>Repeatability</i>	<i>Mean</i>	<i>SD</i>	<i>CV</i>
	<i>g/L</i>	<i>g/L</i>	<i>%</i>
PreciControl CC Multi 1	1.53	0.02	1.4

PreciControl CC Multi 2	2.37	0.03	1.5
Human serum 1	0.249	0.008	3.1
Human serum 2	3.88	0.05	1.2
Human serum 3	2.98	0.04	1.3
Human serum 4	5.12	0.06	1.2
<i>Intermediate precision</i>	Mean	SD	CV
	g/L	g/L	%
PreciControl CC Multi 1	1.53	0.03	1.6
PreciControl CC Multi 2	2.37	0.04	1.7
Human serum 1	0.249	0.010	4.2
Human serum 2	3.88	0.06	1.4
Human serum 3	2.98	0.04	1.4
Human serum 4	5.12	0.07	1.3

**Method comparison****CSF:**

IgA values for human CSF samples obtained on a Roche/Hitachi **cobas c** 501 analyzer (y) were compared with those determined using a nephelometric IgA test (x).

Sample size (n) = 340

Passing/Bablok <sup>18</sup>	Linear regression
$y = 0.925x + 0.0264 \text{ mg/L}$	$y = 0.831x + 0.434 \text{ mg/L}$
$r = 0.963$	$r = 0.983$

The sample concentrations were between 0.466 and 28.1 mg/L.

**Serum/plasma:**

IgA values for human serum and plasma samples obtained on a Roche/Hitachi **cobas c** 501 analyzer (y) were compared with those determined with a nephelometric IgA test (x).

Sample size (n) = 352

Passing/Bablok <sup>18</sup>	Linear regression
$y = 1.09x - 0.112 \text{ g/L}$	$y = 1.07x - 0.0740 \text{ g/L}$
$r = 0.930$	$r = 0.991$

The sample concentrations were between 0.267 and 6.13 g/L.

**References**

- Reiber H, Peter JB. Cerebrospinal fluid analysis: disease-related data patterns and evaluation programs. *J Neurol Sci* 2001;184:101-122.
- Thomas L. Labor und Diagnose. 7. Auflage; TH-Books Verlagsgesellschaft mbH, Frankfurt/Main, 2007;1743-1784.
- Sindic CJ, Delacroix DL, Vaerman JP, et al. Study of IgA in the cerebrospinal fluid of neurological patients with special reference to size, subclass and local production. *J Neuroimmunol* 1984 Dec;7(2-3):65-75.
- Leary SM, McLean BN, Thompson EJ. Local synthesis of IgA in the cerebrospinal fluid of patients with neurological diseases. *J Neurol Sci* 2000;247(8):609-615.
- Woof JM, Mestecky J. Mucosal immunoglobulins. *Immunological Reviews* 2005;206:64-82.
- Occupational Safety and Health Standards: bloodborne pathogens. (29 CFR Part 1910.1030). Fed. Register.
- Directive 2000/54/EC of the European Parliament and Council of 18 September 2000 on the protection of workers from risks related to exposure to biological agents at work.
- WHO Publication: Use of anticoagulants in diagnostic laboratory investigations, WHO/DIL/LAB/99.1 Rev.2:Jan 2002
- Zegers I, Schreiber W, Sheldon J, et al. Certification of proteins in the human serum, ERM-DA470k/IFCC, Report EUR 23431 EN - 2008:1-60 (<https://web.jrc.ec.europa.eu/rmcatalogue/detailsrmcatalogue.do?referenceMaterial=DA470k>).

- Reiber H. The hyperbolic function: a mathematical solution of the protein flux/CSF flow model for blood-CSF barrier function. *J Neurol Sci* 1994;126:243-245.
- Glick MR, Ryder KW, Jackson SA. Graphical Comparisons of Interferences in Clinical Chemistry Instrumentation. *Clin Chem* 1986;32:470-475.
- Breuer J. Report on the Symposium "Drug effects in Clinical Chemistry Methods". *Eur J Clin Chem Clin Biochem* 1996;34:385-386.
- Sonntag O, Scholer A. Drug interference in clinical chemistry: recommendation of drugs and their concentrations to be used in drug interference studies. *Ann Clin Biochem* 2001;38:376-385.
- Attaelmann M, Levinson SS. Understanding and identifying gammopathies. *Clin Chem* 2000;46(8 Pt 2):1230-1238.
- Reiber H, Thompson EJ, Grimsley G, et al. Quality Assurance for Cerebrospinal Fluid Protein Analysis: International Consensus by an Internet-based Group Discussion. *Clin Chem Lab Med* 2003;41:331-337.
- Konsensuswerte der Deutschen Gesellschaft für Laboratoriumsmedizin, der Deutschen Gesellschaft für Klinische Chemie und des Verbandes der Diagnostica-Industrie e.V. (VDGH). *Clin Lab* 1995;41:743-748.
- Lockitch G, Halstead AC, Quigley G, et al. Age- and sex-specific pediatric reference intervals; study design and methods illustrated by measurement of serum proteins with the Behring LN Nephelometer. *Clin Chem* 1988;34:1618-1621.
- Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. *J Clin Chem Clin Biochem* 1988 Nov;26(11):783-790.

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

**Symbols**

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard.

**CONTENT**



Contents of kit

Volume after reconstitution or mixing

COBAS, COBAS C, PRECICONTROL and TINA-QUANT are trademarks of Roche.

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