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RF-II

cobas®

Rheumatoid Factors II

Order information

REF	CONTENT		Analyzer(s) on which **cobas c** pack(s) can be used
20764574 322	Rheumatoid Factors II 100 tests	System-ID 07 6457 4	Roche/Hitachi **cobas c** 311, **cobas c** 501/502
12172828 322	Preciset RF (5 x 1 mL)	Codes 725-729	
03005496 122	RF Control Set (4 x 1 mL)	Code 215 Level I	
		Code 216 Level II	
04489357 190	Diluent NaCl 9 % (50 mL)	System-ID 07 6869 3	

English

System information

For **cobas c** 311/501 analyzers:

RF-II: ACN 017

For **cobas c** 502 analyzer:

RF-II: ACN 8017

Intended use

In vitro test for the quantitative determination of Rheumatoid Factors (RF-II) in human serum and plasma on Roche/Hitachi **cobas c** systems. Measurements may be used as an aid in the diagnosis of rheumatoid arthritis.

Summary[1,2,3,4,5,6,7,8,9,10]

Rheumatoid factors are a heterogeneous group of autoantibodies directed against the antigenic determinants on the Fc-region of IgG molecules. They are important in the diagnosis of rheumatoid arthritis, but can also be found in other inflammatory rheumatic diseases and in various non-rheumatic diseases. They are also found in clinically healthy persons over 60 years of age. Despite these restrictions, the detection of rheumatoid factors is a diagnostic criterion of the American College of Rheumatology for classifying rheumatoid arthritis. The autoantibodies occur in all the immunoglobulin classes, although the usual analytical methods are limited to the detection of rheumatoid factors of the IgM type.

The classic procedure for the quantitation of rheumatoid factors is by agglutination with IgG-sensitized sheep erythrocytes or latex particles. Particular problems of these semiquantitative methods are the poor between-laboratory precision and reproducibility, together with standardization difficulties. For these reasons, new assay methods such as nephelometry, turbidimetry, enzyme-immunoassays and radioimmunoassays have been developed. The Roche RF assay is based on the immunological agglutination principle with enhancement of the reaction by latex.

Test principle[4,5,6]

Immunoturbidimetric assay.

Latex-bound heat-inactivated IgG (antigen) reacts with the RF-antibodies in the sample to form antigen/antibody complexes which, following agglutination, are measured turbidimetrically.

Reagents - working solutions

R1   Glycine buffer: 170 mmol/L, pH 8.0; polyethylene glycol: 0.05 %; bovine serum albumin; stabilizer; preservative

R2   Latex particles coated with human IgG; glycine buffer: 170 mmol/L, pH 7.3; stabilizer; preservative

R1 is in position B and R2 is in position C.

Precautions and warnings

For in vitro diagnostic use.

Exercise the normal precautions required for handling all laboratory reagents.

Disposal of all waste material should be in accordance with local guidelines.

Safety data sheet available for professional user on request.

All human material should be considered potentially infectious. All products derived from human blood are prepared exclusively from the blood of donors tested individually and shown to be free from HBsAg and antibodies to HCV and HIV.

The testing methods applied were FDA-approved or cleared in compliance with the European Directive 98/79/EC, Annex II, List A.

However, as no testing method can rule out the potential risk of infection with absolute certainty, the material should be handled with the same level of care as a patient specimen. In the event of exposure, the directives of the responsible health authorities should be followed.[11,12]

Reagent handling

Ready for use

Mix **cobas c** pack well before placing on the analyzer.

Carefully invert reagent container several times prior to use to ensure that the reagent components are mixed.

Storage and stability

RF-II

Shelf life at 2-8 °C:	See expiration date on **cobas c** pack label.
On-board in use and refrigerated on the analyzer:	8 weeks

Diluent NaCl 9 %

Shelf life at 2-8 °C:	See expiration date on **cobas c** pack label.
On-board in use and refrigerated on the analyzer:	12 weeks

Specimen collection and preparation

For specimen collection and preparation only use suitable tubes or collection containers.

Only the specimens listed below were tested and found acceptable.
Serum
Plasma: Li-heparin and K_2-EDTA plasma

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay.

Stability:[13]	24 hours at 15-25 °C
	3 days at 2-8 °C
	4 weeks at (-15)-(-25) °C (freeze only once)

Materials provided

See "Reagents – working solutions" section for reagents.

Materials required (but not provided)

▪   See "Order information" section

General laboratory equipment

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

The performance of applications not validated by Roche is not warranted and must be defined by the user.

Application for serum and plasma

cobas c 311 test definition

Assay type	2-Point End
Reaction time/Assay points	10 / 7-18
Wavelength (sub/main)	800/570 nm
Reaction direction	Increase

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Rheumatoid Factors II

Unit	IU/mL		
Reagent pipetting		Diluent (H₂O)	
R1	90 µL	–	
R2	30 µL	–	

Sample volumes	Sample	Sample dilution	
		Sample	Diluent (NaCl)
Normal	3 µL	–	–
Decreased	6 µL	15 µL	135 µL
Increased	3 µL	–	–

cobas c 501 test definition

Assay type	2-Point End
Reaction time/Assay points	10 / 12-26
Wavelength (sub/main)	800/570 nm
Reaction direction	Increase
Unit	IU/mL
Reagent pipetting	Diluent (H₂O)
R1	90 µL –
R2	30 µL –

Sample volumes	Sample	Sample dilution	
		Sample	Diluent (NaCl)
Normal	3 µL	–	–
Decreased	6 µL	15 µL	135 µL
Increased	3 µL	–	–

cobas c 502 test definition

Assay type	2-Point End
Reaction time/Assay points	10 / 12-26
Wavelength (sub/main)	800/570 nm
Reaction direction	Increase
Unit	IU/mL
Reagent pipetting	Diluent (H₂O)
R1	90 µL –
R2	30 µL –

Sample volumes	Sample	Sample dilution	
		Sample	Diluent (NaCl)
Normal	3 µL	–	–
Decreased	6 µL	15 µL	135 µL
Increased	6 µL	–	–

Calibration

Calibrators	S1: H₂O
	S2-6: Preciset RF
Calibration mode	RCM
Calibration frequency	Full calibration
	• after 180 days during shelf life
	• after reagent lot change
	• as required following quality control procedures

Traceability: This method has been standardized using the WHO Standard 64/2.

Quality control

For quality control, use control materials as listed in the "Order information" section.

In addition, other suitable control material can be used.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

Calculation

Roche/Hitachi **cobas c** systems automatically calculate the analyte concentration of each sample.

Limitations - interference

Criterion: Recovery within ± 10 % of initial value at an RF concentration of 14 IU/mL.

Icterus:[14] No significant interference up to an I index of 40 for conjugated and 60 for unconjugated bilirubin (approximate conjugated bilirubin concentration: 624 µmol/L or 40 mg/dL) and approximate unconjugated bilirubin concentration: 1026 µmol/L or 60 mg/dL).

Hemolysis:[14] No significant interference up to an H index of 300 (approximate hemoglobin concentration: 186 µmol/L or 300 mg/dL).

Lipemia (Intralipid):[14] No significant interference up to an L index of 2000. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

Drugs: No interference was found at therapeutic concentrations using common drug panels.[15,16]

High dose hook-effect: Using the prozone check, no false result without a flag was observed up to an RF concentration of 6000 IU/mL.

In very rare cases, gammopathy, in particular type IgM (Waldenström's macroglobulinemia), may cause unreliable results.[17]

There is the possibility that other substances and/or factors may interfere with the test and cause unreliable results.

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

ACTION REQUIRED

Special Wash Programming: The use of special wash steps is mandatory when certain test combinations are run together on Roche/Hitachi **cobas c** systems. The latest version of the carry-over evasion list can be found with the NaOHD/SMS/Multiclean/SCCS or the NaOHD/SMS/SmpCln1+2/SCCS Method Sheets. For further instructions refer to the operator's manual. **cobas c** 502 analyzer: All special wash programming necessary for avoiding carry-over is available via the **cobas** link, manual input is not required.

Where required, special wash/carry-over evasion programming must be implemented prior to reporting results with this test.

Limits and ranges
Measuring range

10-130 IU/mL

Determine samples having higher concentrations via the rerun function. Dilution of samples via the rerun function is a 1:5 dilution. Results from samples diluted using the rerun function are automatically multiplied by a factor of 5.

Lower limits of measurement

Lower detection limit of the test

10 IU/mL

The lower detection limit represents the lowest measurable analyte level that can be distinguished from zero. It is calculated as the value lying three standard deviations above that of the lowest standard (standard 1 + 3 SD, repeatability, n = 21).

Expected values[18]

< 14 IU/mL

This value is based on serum samples from 525 test subjects.

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

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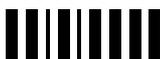
Rheumatoid Factors II

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using human samples and controls in an internal protocol with repeatability (n = 21) and intermediate precision (3 aliquots per run, 1 run per day, 21 days). The following results were obtained:

Repeatability	Mean IU/mL	SD IU/mL	CV %
RF Control level 1	23.7	0.2	0.8
RF Control level 2	53.0	0.5	0.9
Human serum 1	19.5	0.3	1.6
Human serum 2	27.5	0.3	1.1

Intermediate precision	Mean IU/mL	SD IU/mL	CV %
RF Control level 1	23.2	0.3	1.4
RF Control level 2	51.4	0.8	1.5
Human serum 3	19.3	0.3	1.6
Human serum 4	26.1	0.5	1.8

Method comparison

RF values for human serum and plasma samples obtained on a Roche/Hitachi **cobas c** 501 analyzer (y) were compared with those determined using the corresponding reagent on a Roche/Hitachi 917 analyzer (x).

Sample size (n) = 70

Passing/Bablok[19]	Linear regression
y = 1.000x - 1.20 IU/mL	y = 0.999x - 1.39 IU/mL
τ = 0.959	r = 0.998

The sample concentrations were between 10.8 and 114 IU/mL.

References

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2 Arnett FC, Edworthy SM, Bloch DA, et al. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. Arthritis Rheum 1988;31:315-324.

3 Bartfield H. Distribution of rheumatoid factor activity in non-rheumatoid states. Ann NY Acad Sci 1969;16:30-40.

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5 Moore TL, Dorner RN. Rheumatoid factors. Clin Biochem 1993;26:75-84.

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11 Occupational Safety and Health Standards: bloodborne pathogens. (29 CFR Part 1910.1030). Fed. Register.

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15 Breuer J. Report on the Symposium "Drug effects in Clinical Chemistry Methods". Eur J Clin Chem Clin Biochem 1996;34:385-386.

16 Sonntag O, Scholer A. Drug interference in clinical chemistry: recommendation of drugs and their concentrations to be used in drug interference studies. Ann Clin Biochem 2001;38:376-385.

17 Bakker AJ, Mücke M. Gammopathy interference in clinical chemistry assays: mechanisms, detection and prevention. Clin Chem Lab Med 2007;45(9):1240-1243.

18 Data on file at Roche Diagnostics.

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A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard.

CONTENT	Contents of kit
⟶	Volume after reconstitution or mixing

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