

Reticulum II Staining Kit

REF 860-024

05279399001

IVD  75

INTENDED USE

Reticulum II Staining Kit is intended for use as a qualitative histologic stain to demonstrate reticular fibers in formalin-fixed, paraffin-embedded tissue.

This product should be interpreted by a qualified pathologist in conjunction with histological examination, relevant clinical information, and proper controls.

This product is intended for *in vitro* diagnostic (IVD) use.

SUMMARY AND EXPLANATION

The purpose of this stain is primarily to aid in the differential diagnosis of certain tumor types. In certain tumors, reticulum is located in a characteristic position in relation to the actual tumor cells. Reticulum stain can be used to show disease states in organs such as the liver, spleen and kidney by demonstrating reticular patterns not normal to the organ. In normal liver, the fibers are well defined strands, but in necrotic or cirrhotic liver, the fibers have discontinuous patterns.¹

Reticulum II Staining Kit is a modification of Gordon and Sweets Stain.² Oxidizer, with potassium permanganate, oxidizes the tissue to enhance staining of reticular fibers. Decolorizer, with oxalic acid, removes excess potassium permanganate. Sensitizer, with ferric ammonium sulfate, is added to form a metal organic compound. The metal organic compound is replaced by the silver in Reticulum II Silver A. Reducer is applied to develop the deposited silver into visible silver. Toner reagent contains gold chloride for better contrast and clarity. Fixer, with thiosulfate, stops the reaction and removes any unreacted silver from the section. Nuclear Fast Red Counterstain is applied to provide contrasting background.

PRINCIPLE OF THE PROCEDURE

This kit is optimized for use on VENTANA BenchMark Special Stains automated slide stainers and VENTANA NexES Special Stains automated slide stainers. The reagents are applied to tissue on microscope slides and mixed over the entire specimen. The staining reaction is based on the affinity of silver for oxidized glycoproteins.

MATERIALS AND METHODS

Reagents Provided

The reagent vials are supplied in barcode labeled carriers to insert into the reagent tray of the automated slide stainer. Each kit contains sufficient reagent for 75 tests:

One 27 mL vial of Oxidizer contains less than 1% potassium permanganate.

One 22 mL vial of Decolorizer contains less than 1% oxalic acid.

One 27 mL vial of Sensitizer contains 2% ferric ammonium sulfate.

One 22 mL vial of Reticulum II Silver A contains less than 1.5% silver carbonate.

One 22 mL vial of Reticulum II Reducer contains 0.4% formaldehyde.

One 22 mL vial of Toner contains less than 1% gold chloride.

One 22 mL vial of Fixer II contains 2% sodium thiosulfate.

One 22 mL vial of Nuclear Fast Red Counterstain contains less than or equal to 0.2% Nuclear Fast Red and less than or equal to 5.0% aluminum sulfate.

8 vial inserts with sipping straws.

Reconstitution, Mixing, Dilution, Titration

No reconstitution, mixing, dilution, or titration of kit reagents is required. Further dilution of any of the reagents may result in unsatisfactory staining quality. The user must validate any such change.

The reagents in this kit have been optimally diluted for use on BenchMark Special Stains automated slide stainers and NexES Special Stains automated slide stainers. These reagents may not be optimal for manual procedures or for use on other instruments.

Materials and Reagents Needed But Not Provided

- Control tissue
- Microscope slides (positively charged)
- Drying oven capable of maintaining a temperature of 70°C ± 5°C
- BenchMark Special Stains automated slide stainer or NexES Special Stains automated slide stainer.
- Bulk reagents for the BenchMark Special Stains automated slide stainer:
 - BenchMark Special Stains Deparaffinization Solution (10X)
 - BenchMark Special Stains Liquid Coverslip
 - BenchMark Special Stains Wash Solution (10X)
- Bulk reagents for the NexES Special Stains automated slide stainer:
 - Liquid Coverslip (Low Temperature)
 - Special Stains Wash (10X)
- Xylene or other clearing reagent (Histological grade)
- Reagent alcohol or ethanol (Histological grade)
- Deionized or distilled water
- Synthetic mounting media
- Coverslip

Storage and Handling

The Reticulum II Staining Kit should be stored at 2-8°C. See vial label for proper storage conditions. Refrigerated kit components should be brought to room temperature prior to use.

When properly stored, reagents are stable until the expiration date that is printed on the vial label. There are no obvious signs to indicate instability of these reagents; therefore, controls should be run simultaneously with unknown specimens. If positive control material shows a decrease in staining it could indicate reagent instability and your local support representative should be contacted immediately.

Specimen Collection and Preparation for Analysis

Ventana recommends specimen collection and storage be performed according to CLSI document M29-T2.³ The recommended tissue fixative is 10% neutral buffered formalin.²

- Cut sections, usually 3 to 5 µm, and pick up the sections on glass slides.
- You can either bake the slides on the BenchMark Special Stains instrument or use an alternative method off the instrument.
 - If you choose to bake the slides on the BenchMark Special Stains instrument, proceed to step 3.
 - Off the instrument, bake the slides for at least 30 minutes at approximately 70°C. Allow to cool.
- Print appropriate barcode label(s).
- Apply barcode labels to the frosted end of the slides prior to deparaffinization (see the instrument Operator's Manual for correct application of labels).
- Refer to the Instructions for Use section for the recommended protocol for the BenchMark Special Stains automated slide stainer and NexES Special Stains automated slide stainer.

WARNINGS AND PRECAUTIONS

- For *in vitro* diagnostic (IVD) use.
- For professional use only.
- Consult local and/or state authorities with regard to recommended method of disposal.
- Materials of human or animal origin should be handled as biohazardous materials and disposed of with proper precautions. Use universal precautions when handling and disposing of specimens.
- Avoid microbial contamination of reagents. Contamination could produce erroneous results.
- These reagents may cause irritation. Avoid contact with eyes and mucous membranes. If reagent contacts these areas, rinse with copious amounts of water.
- Oxidizer is considered harmful to aquatic organisms, may cause long-term adverse effects in the aquatic environment.
- Reticulum II Reducer is highly flammable and an irritant. May cause sensitization by skin contact.
- Fixer II is highly flammable.

10. Nuclear Fast Red Counterstain is considered an irritant to eyes.
11. For supplementary safety information, refer to the product Safety Data Sheet and the Symbol and Hazard Guide located at www.ventana.com.

INSTRUCTIONS FOR USE

Prepare Reagent Vial

Before first use, a vial insert and sipping straw must be placed in the reagent vial.

1. Remove the shipping cap from the vial and place the insert and straw into the vial. The insert and sipping straw should be left in the vial, once the vial has been opened.
2. Place the soft cap into the slot on the reagent holder when the reagent is in use.
3. Use the soft cap to cover the reagent vial when reagent is not in use.

Staining Procedure

1. Load reagents and slides onto the instrument.
2. Perform the staining run according to the recommended protocol (see Table 1 or Table 2) and the instructions in the manual.
3. When the run is complete, remove the slides from the instrument.

Recommended Protocol

The parameters for the automated procedures can be displayed, printed and edited according to the procedure in the instrument Operator's Manual.

The following procedures allow flexibility to accommodate varying tissue thickness, size and user preference. Trial runs using the protocols are suggested to adjust staining to the user's preference.

In the protocol in Table 1, the protocol selections are necessary to enable deparaffinization and baking for the BenchMark Special Stains instrument.

Table 1. Recommended Staining Protocol for Reticulum II Staining Kit on a BenchMark Special Stains Automated Slide Stainer.

| Staining Procedure: S Reticulum II version 2 | |
|--|---|
| Protocol Step | Method |
| Deparaffinization | Select deparaffinization to automate paraffin removal. |
| Baking | Select temperature and incubation time to enable baking. |
| Temperature | Select an incubation temperature from 37°C to 60°C. <ul style="list-style-type: none"> • 37°C, lighter staining of fibers • 60°C, darker staining of fibers Ventana recommends starting at 40°C. Adjust temperature in 1°C increments until desired staining intensity of fibers is achieved. |
| Oxidizer | Choose an incubation time from 4-20 minutes for each parameter. Ventana recommends starting with the following incubation times: <ul style="list-style-type: none"> • Oxidizer 4 minutes • Decolorizer 8 minutes • Sensitizer 8 minutes Ventana recommends Decolorizer incubation time remains at least 2 times the incubation time of Oxidizer. If Oxidizer incubation time is increased beyond the recommended starting time of 4 minutes, increase the Decolorizer incubation time accordingly. |
| Decolorizer | |
| Sensitizer | |
| Optimize Silver Intensity (Reticulum II Silver A) | Select an incubation time from 8 to 20 minutes: <ul style="list-style-type: none"> • 8 minutes, lighter staining of fibers • 20 minutes, darker staining of fibers Ventana recommends starting with a 16 minute incubation time. If Sensitizer incubation time is increased beyond the recommended starting time of 8 minutes, increase the Reticulum II Silver A incubation time. |
| Optimize Counterstain Intensity (Nuclear Fast Red Counterstain) | Select an incubation time from 4 to 16 minutes: <ul style="list-style-type: none"> • 4 minutes, lighter counterstain • 16 minutes, darker counterstain Ventana recommends starting with a 4 minute incubation time. |
| No Counterstain | If Nuclear Fast Red Counterstain is not required, select No Counterstain. |

Table 2. Recommended Staining Protocol for Reticulum II Staining Kit on a NexES Special Stains Automated Slide Stainer.

| Procedure Type | Method |
|--------------------------------|---|
| Deparaffinization | <ol style="list-style-type: none"> 1. Deparaffinize and hydrate slides following recommended procedures within the laboratory. 2. Place tissue slides in Special Stains Wash Solution (1X) until loading the instrument. It is recommended that slides be loaded on the instrument within one hour. If not possible the slides may be left in deionized or distilled water for up to 24 hours as long as the tissue is placed in the refrigerator and not allowed to dry. |
| Options | |
| Temperature | 37°C or 60°C |
| Time | 4 to 12 minutes for Reticulum II Silver A |
| Counterstain | Light or Dark |
| Pre-Programmed Protocol | |
| 37°C protocols | Have protocol numbers starting with 3XX |
| 60°C protocols | Have protocol numbers starting with 6XX |

Recommended Post-Instrument Processing

1. Rinse slides in three changes of 95% alcohol to remove the coverslip solution.
2. Dehydrate, clear, and coverslip with permanent mounting media.

QUALITY CONTROL PROCEDURES

An example of a positive control material would be formalin-fixed, paraffin-embedded human tissue with defined reticular fibers (liver, spleen, kidney, or lymph nodes).² Control tissue should be fresh autopsy, biopsy, or surgical specimen prepared or fixed as soon as possible in a manner identical to test sections. Such tissues should monitor all steps of the analysis, from tissue preparation through staining.

Use of a tissue section fixed or processed differently from the test specimen provides control for all reagents and method steps except fixation and tissue processing. The cellular components of other tissue elements may serve as the negative control.

The control tissue must be tested with each run.

The Reticulum II Staining Kit is tested upon manufacture to show that reticular fibers stain black against a pink to red background.

Known positive tissue controls should only be utilized for monitoring the correct performance of processed tissues and test reagents, not as an aid in formulating a specific diagnosis of patient samples.

If the positive tissue components fail to demonstrate positive staining, results with the test specimens should be considered invalid. If the negative components demonstrate positive staining, results with patient specimens should also be considered invalid.

Unexplained discrepancies in control results should be referred to the local support representative immediately. If quality control results do not meet specifications, patient results are invalid. The cause must be identified and corrected, and the patient samples repeated.

LIMITATIONS

General Limitations

1. Histological staining is a multiple step diagnostic process that requires specialized training in the selection of the appropriate reagents, tissue selections, fixation, processing, preparation of the slide, and interpretation of the staining results.
2. Tissue staining is dependent on the handling and processing of the tissue prior to staining. Differences in tissue processing and technical procedures may produce significant variability of results necessitating regular performance of controls (see the Quality Control Procedures section). Improper fixation, freezing, thawing, washing, drying, heating, sectioning, or contamination with other tissues or fluids may produce artifacts or false negative results.

3. The clinical interpretation of any positive staining, or its absence, must be evaluated within the context of clinical history, morphology and other histopathological criteria. It is the responsibility of a qualified pathologist to be familiar with the special stain and methods used to produce the slide. Staining must be performed in a certified licensed laboratory under the supervision of a pathologist who is responsible for reviewing the stained slides and assuring the adequacy of positive and negative controls.

PERFORMANCE CHARACTERISTICS

BenchMark Special Stains Automated Slide Stainer

1. Instrument-to-Instrument: 100 slides of liver tissue were tested across 5 different BenchMark Special Stains instruments with the Reticulum II Staining Kit. The slides were evaluated for staining with pass or fail criteria. The results demonstrated no significant difference in staining intensity among the slides.
2. Run-to-Run: 100 slides of liver tissue were tested across 5 BenchMark Special Stains instruments on 5 non-consecutive days with the Reticulum II Staining Kit. The slides were evaluated for staining with pass or fail criteria. The results demonstrated no significant difference in staining intensity among slides.
3. Compatibility with Reticulum II Staining Kit was tested with other selected special stains reagents. No significant adverse chemical interactions were observed.

NexES Special Stains Automated Slide Stainer

1. Intra-run reproducibility of Reticulum II Staining Kit was determined by staining slides from 8 different liver and hepatocellular carcinoma tissue blocks for a total of 100 slides. The slides were evaluated for staining performance (overall coverage and quality of the stain), diagnostic utility (the disease state can be assessed from the stain), background and counterstain with pass or fail criteria. The results demonstrated that 100/100 slides performed per the acceptance criteria.
2. Inter-run reproducibility of staining with Reticulum II Staining Kit was determined by staining 20 slides per run across five instruments (1 run per instrument). The slides were evaluated for staining performance, diagnostic utility; background and counterstain with pass or fail criteria. The results demonstrated no significant difference in staining intensity among slides.

TROUBLESHOOTING

1. Section thickness may affect quality and intensity of staining. If staining is inappropriate, contact your local support representative for assistance.
2. Necrotic or autolyzed tissue may exhibit nonspecific staining.
3. If the positive control does not stain appropriately, check to ensure the slide has the correct barcode label and it is applied correctly. If the label is correct, but no staining or unexpected staining occurs, contact your local support representative.

REFERENCES

1. Sheehan DC, Hrapchak BB. Theory and Practice of Histotechnology, 2nd edition. St. Louis, MO: C.V. Mosby Company; 1980:181-188.
2. Carson F, Hladik C. Histotechnology: A Self Instructional Text, 3rd edition. Hong Kong: American Society for Clinical Pathology Press; 2009.
3. Clinical and Laboratory Standards Institute (CLSI). CLSI Web site. <http://www.clsi.org/>. Accessed November 3, 2011.

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CONTACT INFORMATION

Ventana Medical Systems, Inc.
1910 E. Innovation Park Drive
Tucson, Arizona 85755
USA

+1 520 887 2155
+1 800 227 2155 (USA)



www.ventana.com



Roche Diagnostics GmbH
Sandhofer Strasse 116
D-68305 Mannheim
Germany