

• Indicates **cobas c** systems on which reagents can be used**Order information**

ONLINE TDM Theophylline

100 Tests	Cat. No. 04491025 190	System-ID 07 6927 4
Preciset TDM I calibrators	Cat. No. 03375790 190	
CAL A-F	1 x 5 mL	Codes 691-696
Diluent	1 x 10 mL	
TDM Control Set	Cat. No. 04521536 190	
Level I	2 x 5 mL	Code 310
Level II	2 x 5 mL	Code 311
Level III	2 x 5 mL	Code 312

Roche/Hitachi **cobas c** systems**cobas c** 501/502

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English**System information**For **cobas c** 501 analyzer:**THEO2**: ACN 415For **cobas c** 502 analyzer:**THEO2**: ACN 8415**Intended use**

In vitro test for the quantitative determination of theophylline in serum and plasma on Roche/Hitachi **cobas c** systems.

Summary

Theophylline (1,3-dimethylxanthine), a bronchodilator, is widely used to treat patients with asthma, apnea (temporary asphyxia), and other obstructive lung diseases. Monitoring of theophylline concentrations in serum is essential, since individuals can vary in their rates of theophylline clearance,^{1,2} and severe toxicity has been observed without prior occurrence of minor side effects.³ Moreover, several factors can alter theophylline elimination. Theophylline elimination is slowed in obese patients, patients with hepatic disease, and in those on a high carbohydrate, low protein diet. Premature infants have very low rates of theophylline elimination.⁴ Conversely, theophylline elimination is more rapid among cigarette smokers.⁵ In combination with other clinical data, monitoring serum theophylline levels may provide the physician with useful information to aid in adjusting patient dosage to achieve optimal therapeutic effect while avoiding drug toxicity.

Test principle

The assay is based on the kinetic interaction of microparticles in a solution (KIMS). Theophylline antibody is covalently coupled to microparticles and the drug derivative is linked to a macromolecule. The kinetic interaction of microparticles in solutions is induced by binding of drug-conjugate to the antibody on the microparticles and is inhibited by the presence of theophylline in the sample. A competitive reaction takes place between the drug conjugate and theophylline in the serum sample for binding to the theophylline antibody on the microparticles. The resulting kinetic interaction of microparticles is indirectly proportional to the amount of drug present in the sample.

Reagents - working solutions

- R1** Theophylline conjugate; piperazine-N,N'-bis (ethanesulfonic acid) (PIPES) buffer, pH 7.2; preservative
- R2** Anti-theophylline antibody (mouse monoclonal); latex microparticle; 3-(N-morpholino) propane sulfonic acid (MOPS) buffer, pH 7.5; stabilizer; preservative
- R1 is in position B and R2 is in position C.

Precautions and warnings

For in vitro diagnostic use.

Exercise the normal precautions required for handling all laboratory reagents. Safety data sheet available for professional user on request.

Disposal of all waste material should be in accordance with local guidelines.

Reagent handling

Ready for use. Mix reagents by gentle inversion numerous times before placing on-board the analyzer.

Storage and stability

Shelf life at 2 to 8 °C:

See expiration date on

cobas c pack label

On-board in use and refrigerated on the analyzer: 12 weeks

Do not freeze.**Specimen collection and preparation**

For specimen collection and preparation, only use suitable tubes or collection containers.

Only the specimens listed below were tested and found acceptable.

Serum: Collect serum using standard sampling tubes.

Plasma: K₂- or K₃-EDTA, sodium citrate, or sodium, lithium or ammonium heparin plasma.

Stability:⁶

1 week capped at 2-8 °C

60 days capped at -20 °C

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer. Centrifuge samples containing precipitates before performing the assay. Specimens should not be repeatedly frozen and thawed. Invert thawed specimens several times prior to testing. Usual sampling time varies dependent upon desired measurement of peak or trough values.⁷

Materials provided

See "Reagents - working solutions" section for reagents.

Materials required (but not provided)

See "Order information" section

General laboratory equipment

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

The performance of applications not validated by Roche is not warranted and must be defined by the user.

Application for serum and plasma

Deselect Automatic Rerun for this application in the Utility menu, Application screen, Range tab.

cobas c 501/502 test definition

Assay type	2 Point End	
Reaction time / Assay points	10 / 15-49	
Wavelength (sub/main)	800 / 600 nm	
Reaction direction	Increase	
Unit	µg/mL	
Reagent pipetting		Diluent (H ₂ O)
R1	97 µL	–
R2	92 µL	–

Theophylline

Sample volumes	Sample	Sample dilution	
		Sample	Diluent (NaCl)
Normal	2.0 µL	–	–
Decreased	2.0 µL	–	–
Increased	2.0 µL	–	–

Calibration

Calibrators	S1-6: Preciset TDM I calibrators
Calibration mode	RCM
Calibration frequency	6-point calibration <ul style="list-style-type: none"> - after cobas c pack change - every 6 weeks - as required following quality control procedures

Traceability: This method has been standardized against USP reference standards. The calibrators are prepared to contain known quantities of theophylline in normal human serum.

Quality Control

For quality control use the control material as listed in the "Materials required" section. In addition, other suitable control material can be used. The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

Calculation

Roche/Hitachi **cobas c** systems automatically calculate the analyte concentration of each sample.

Conversion factor:⁸ µg/mL x 5.55 = µmol/L

Limitations - interference

Criterion: Recovery within ± 10 % of initial value at theophylline levels of approximately 5 and 15 µg/mL (27.8 and 83.3 µmol/L).

Serum/Plasma

Icterus:⁹ No significant interference up to an I index of 50 (approximate conjugated and unconjugated bilirubin concentration: 50 mg/dL or 855 µmol/L).

Hemolysis:⁹ No significant interference up to an H index of 1000 (approximate hemoglobin concentration: 1000 mg/dL or 621 µmol/L).

Lipemia (Intralipid):⁹ No significant interference up to an L index of 300. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

No significant interference from triglycerides up to 1000 mg/dL (11.3 mmol/L).

Rheumatoid factors: No significant interference from rheumatoid factors up to 100 IU/mL.

Total protein: No interference from total protein up to 12 g/dL.

Theobromine: No significant interference up to 20 µg/mL theobromine. Concentrations above this toxic level may result in negative bias of > 10 %.

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

ACTION REQUIRED

Special Wash Programming: The use of special wash steps is mandatory when certain test combinations are run together on Roche/Hitachi **cobas c** systems. The latest version of the Carry-over evasion list can be found with the NaOHD/SMS/Multiclean/SCCS or the NaOHD/SMS/SmpCln1 + 2/SCCS Method Sheets. For further instructions refer to the operator's manual.

cobas c 502 analyzer: All special wash programming necessary for avoiding carry-over is available via **cobas** link, manual input is not required.

Where required, special wash/carry-over evasion programming must be implemented prior to reporting results with this test.

Limits and ranges

Measuring range

0.8-40.0 µg/mL (4.4-222 µmol/L)

Manually dilute samples having higher concentrations with Preciset TDM I diluent (0 µg/mL) (1 + 1) and reassay. Multiply the result by 2 to obtain the specimen value.

Lower limits of measurement

Lower detection limit of the test

0.8 µg/mL (4.4 µmol/L)

The lower detection limit represents the lowest measurable analyte level that can be distinguished from zero. It is calculated as the value lying two standard deviations above that of the 0 µg/mL calibrator (standard 1 + 2 SD, repeatability, n = 21).

Expected values

Various methodologies have been used to evaluate theophylline preparations and routes of administration,¹⁰ to study pharmacokinetics of the drug,¹¹ and to define the relationship between serum concentration and the drug's therapeutic and toxic effects.¹² For most patients, the range of 10 to 20 µg/mL (55.5 to 111 µmol/L) suppresses chronic asthmatic symptoms.^{13,14,15,16} Wide discrepancies between drug dosage and serum concentrations were observed among patients receiving identical doses.¹³ A major factor accounting for the variability is individual variation in the rate of theophylline metabolism and elimination.

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data¹⁷

Representative performance data on a Roche/Hitachi analyzer are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using human samples and controls in a modified NCCLS EP5-T2 protocol (repeatability* n = 63, intermediate precision** n = 63). The following results were obtained on a Roche/Hitachi **cobas c 501** analyzer.

Serum/Plasma

Repeatability*	Mean		SD		CV %
	µg/mL	µmol/L	µg/mL	µmol/L	
Control 1	4.25	23.6	0.07	0.4	1.7
Control 2	14.3	79.4	0.2	1.1	1.3
Control 3	34.1	189	0.4	2	1.2
HS 1	5.78	32.1	0.08	0.4	1.4
HS 2	20.0	111	0.3	2	1.4

Intermediate precision**	Mean		SD		CV %
	µg/mL	µmol/L	µg/mL	µmol/L	
Control 1	4.25	23.6	0.12	0.7	2.8
Control 2	14.3	79.4	0.2	1.1	1.7
Control 3	34.1	189	0.6	3	1.9
HS 1	5.78	32.1	0.12	0.7	2.1
HS 2	20.0	111	0.4	2	1.8

* repeatability = within-run precision

** intermediate precision = total precision / between run precision / between day precision

Method comparison

Serum/plasma

Theophylline values for human serum and plasma samples obtained on a Roche/Hitachi **cobas c 501** analyzer (y) were compared with those determined using the corresponding reagent on a Roche/Hitachi 917 analyzer (x) and on a COBAS INTEGRA 800 analyzer (x).

Roche/Hitachi 917 analyzer	Sample size (n) = 72
Passing/Bablok ¹⁸	Linear regression
y = 0.975x + 0.136 µg/mL	y = 0.982x + 0.032 µg/mL
τ = 0.985	r = 0.999

The sample concentrations were between 3.98 and 39.0 µg/mL (22.1 and 217 µmol/L).

THEO2

Theophylline

COBAS INTEGRA 800 analyzer

Passing/Bablok¹⁸

$y = 1.017x + 0.091 \mu\text{g/mL}$

$\tau = 0.981$

Sample size (n) = 72

Linear regression

$y = 1.013x + 0.143 \mu\text{g/mL}$

$r = 0.999$

The sample concentrations were between 3.71 and 39.0 $\mu\text{g/mL}$ (20.6 and 217 $\mu\text{mol/L}$).

Analytical specificity

The following compounds were tested for cross-reactivity.

Compound	Concentration Tested ($\mu\text{g/mL}$)	% Cross-reactivity
Aminophylline	15	79.6
8-Chlorotheophylline	200	5.97
1,7-Dimethylxanthine	150	5.24
3-Methylxanthine	150	2.73
Ephedrine	12	1.00
Acetaminophen	200	< 1.0
Allopurinol	50	< 1.0
Caffeine	150	< 1.0
Dihydroxypropyl theophylline	200	< 1.0
Diphenhydramine	10	< 1.0
Epinephrine	16	< 1.0
β -Hydroxyethyl theophylline	200	< 1.0
7- β -Hydroxypropyl theophylline	200	< 1.0
Hypoxanthine	150	< 1.0
Isoproterenol	50	< 1.0
1-Methyluric acid	400	< 1.0
Phenobarbital	200	< 1.0
Phenylbutazone	400	< 1.0
Uric acid	210	< 1.0
1,3-Dimethyluric acid	700	< 0.1
Phenytoin	200	< 0.1

Tests were performed on 15 drugs. No significant interference with the assay was found.

Acetaminophen	Doxycycline (Tetracycline)
Acetyl cysteine	Ibuprofen
Acetylsalicylic acid	Levodopa
Ampicillin-Na	Methyldopa + 1.5 H ₂ O
Ascorbic acid	Metronidazole
Ca-Dobesilate	Phenylbutazone
Cefoxitin	Rifampicin
Cyclosporine	

References

- Piafsky KM, Ogilvie RI. Drug therapy. Dosage of theophylline in bronchial asthma. *N Engl J Med* 1975;292:1218-1222.
- Leung P, Kalisker A, Bell TD. Variation in theophylline clearance rate with time in chronic childhood asthma. *J Allergy Clin Immunol* 1977;59:440-444.
- Zwillich CW, Sutton FD, Neff TA, et al. Theophylline-induced seizures in adults. Correlation with serum concentrations. *Ann Intern Med* 1975;82:784-787.
- Ogilvie RI. Clinical pharmacokinetics of theophylline. *Clinical Pharmacokinetics* 1978;3:267-293.
- Hendeles L, Weinberger MM. Theophylline therapeutic use and serum concentration monitoring. In: Taylor WJ, Finn AL, eds. *Individualizing Drug Therapy: Practical Applications of Drug Monitoring*, I. New York, NY: Gross Townsend Frank Inc 1981:31-66.
- Committee on patient preparation and specimen handling. *Clinical Laboratory Handbook for Patient Preparation and Specimen Handling*. Fascicle IV. Slokie, IL: College of American Pathologists; 1985.
- Jacobs DS, Kasten BL Jr, Demott WR, et al. *Laboratory Test Handbook*. Stow, OH: Lexi-Comp 1990;819.

- Tietz NW. *Clinical Guide to Laboratory Tests*. 3rd ed. Philadelphia, Pa: WB Saunders Co 1995:878.
- Glick MR, Ryder KW, Jackson SA. Graphical Comparisons of Interferences in Clinical Chemistry Instrumentation. *Clin Chem* 1986;32:470-475.
- Truitt EG Jr, McKusick VA, Krantz C. Theophylline blood levels after oral, rectal, and intravenous administration and correlation with diuretic action. *J Pharmacol Exp Ther* 1958;10:310-315.
- Mitenko PA, Ogilvie RI. Pharmacokinetics of intravenous theophylline. *Clin Pharm Therapeutics* 1973;14:509-513.
- Turner-Warwick M. Study of theophylline plasma levels after oral administration of theophylline compounds. *Br Med J* 1957;25.
- Jackson FR, Garrido R, Silverman HI, et al. Blood levels following oral administration of theophylline preparations. *Ann Allergy* 1973;31:413-419.
- Jenne JW, Wyze E, Rood FS, et al. Pharmacokinetics of theophylline: Application to adjustment of the clinical use of aminophylline. *Clin Pharmacol Ther* 1972;13:349-360.
- Weinberger MM, Bronsky EA. Evaluation of oral bronchodilator therapy in asthmatic children. *J Pediatr* 1974;84:421-427.
- Weinberger MM, Riegelman S. Rational use of theophylline for bronchodilation. *N Engl J Med* 1974;291:151-153.
- Data on file at Roche Diagnostics.
- Passing H, Bablok W, Bender R, et al. A General Regression Procedure for Method Transformation. *J Clin Chem Clin Biochem* 1988 Nov;26(11):783-790.

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

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